

Weed Management in Southern Bareroot Hardwood Nurseries

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Abstract

Managers in the southern United States rely on chemicals and non-chemical methods of weed control. Chemical treatments include fumigation with methyl bromide and chloropicrin in combination with selective herbicides. At nurseries where methyl bromide is not used, managers may choose to increase reliance on herbicides and sanitation practices. Although several herbicides are registered for use on hardwoods, some formulations can injure seedlings if improperly applied to seedbeds. Most grasses can be effectively controlled with selective herbicides and many small seeded broadleaf weeds can be suppressed when preemergence herbicides are applied before germination of weeds. Several preemergence herbicides may be legally applied either at time of sowing or after seedlings are established. Several nursery managers have fabricated shielded herbicide sprayers to apply herbicides between drills to certain difficult to control weeds. The most effective programs include a rigorous sanitation program and judicious use of efficacious herbicides.

Introduction

The hardwood nursery manager's primary objective is to produce morphologically-improved stock as economically as possible. Weeds can be a major obstacle to this goal since they compete with seedlings for light, water, and nutrients. In addition, handweeder often pull up seedlings while weeding, reducing revenue from seedling sales. In some cases, weed populations will stunt seedlings and will cause large variations in seedling size at lifting.

To maintain a relatively weed-free nursery, most hardwood nursery managers implement a comprehensive year-round weed control program. In the past, some hardwood seedbeds required more than 1,600 hours of handweeding (Abrahamson 1987). Today, many managers use herbicides as part of an integrated weed management (IWM) program (Walker and Buchanan 1982) which includes sanitation, soil fumigation, and herbicide applications to keep weed populations low and minimize handweeding. As a result, several hardwood nurseries now require less than 25 hours of handweeding per acre.

Weed Identification

To achieve good weed control, weed species must be accurately identified, especially when troublesome species are present. For example, some herbicides will suppress yellow nutsedge (*Cyperus esculentus* L.) but have little effect on purple nutsedge (*Cyperus rotundus* L.) even though the two species appear similar. Several online sites are available for identifying common weeds and extension weed specialists should be able to identify rare species. Some of the more common weeds in southern nurseries, with their scientific names, are listed in Table 1.

Sanitation

Preventing weeds from going to seed in the nursery is an important sanitation practice since weed populations in future years greatly depend upon the number of seed produced during the current season. If one yellow nutsedge plant is allowed to mature, it can produce more than 2,400 seeds. A mature purslane plant (*Portulaca oleracea* L.) can produce over 52,000 seeds and a single redroot pigweed (*Amaranthus retroflexus* L.) can produce 117,000 seeds or more (Stevens 1932). The importance of preventing a single weed from maturing and producing seed in the nursery cannot be overemphasized. A severe infestation of nutsedge can quickly result from failure to control even a single plant. For example, one tuber of purple nutsedge produced 1,168 plants and 2,324 tubers after 6 months (Ishii et al. 1971). Weeds must be prevented from going to seed not only in the seedbeds, but also on the riserlines, fencerows, cover crop areas, and fallow areas (Wichman 1982).

Irrigation Water

Irrigation water can be a major source of introduced weeds when the water is from a lake, pond, or river. The use of screens at the intake pipe can help filter out large-seeded weeds. Although the screens may require frequent cleaning, it is easier to remove the weed seeds from the screens than to remove weeds from seedbeds. When irrigating from ponds, it is best to keep the pond edges free of weeds. When installing a new nursery, a deep well is preferred over surface water sources.

Cover-Crop Seeds

Sowing weed seeds along with cover-crop seeds can be prevented by always using certified seed. At one nursery, the use of cheap, uncertified seed resulted in a large infestation of morning glory (*Ipomoea* spp.). Regulations require certified seed to be free of primary noxious weeds and to contain only small amounts of common weeds. The percentage of common weeds must be shown on the certification tag. It is best to buy seeds with the lowest percentage of common weeds.

Machinery

Weed seeds, rhizomes, and tubers are easily introduced by machinery. Frequent washings reduce the amount of weeds introduced by soil carried on tillage equipment, tractors, and vehicle tires. Weed seeds are often spread by combines during the harvest of cover crops. For this reason, it is better to leave cover crops unharvested unless combines are carefully cleaned before and after use.

Some weeds spread slowly by vegetative means alone. For example, nutsedge would spread less than 10 ft (3 m) per year without help from nursery workers and their cultivation equipment (Klingman and Ashton 1975). For this reason, special efforts should be made to avoid spreading nutsedge. Infested seedbeds can be mapped in the summer to help identify areas in which to avoid soil movement (thus spreading nuts) in the winter after lifting. Nutsedge-free areas should be lifted first to avoid the spreading of tubers to non-infested fields. Time taken to prevent mechanical dissemination of nutsedge tubers will be repaid several-fold in the ease of eliminating nutsedge from a nursery.

Wind

Wind will constantly introduce weed seeds, but the impact may be reduced by planting windbreaks between the nursery and adjacent weed sources, such as farmlands. Windbreaks will also help to protect the nursery from high winds that blow mulch off beds, blow plastic off fumigated soil, and cause excessive drying of the beds.

Mulches

In the past, the use of straw mulches after sowing was a major source of introduced weeds (Mullin 1965; Bland 1973; South 1976). For example, at some nurseries, pine straw mulch increased time spent on handweeding by 105 to 200 hours per acre (Bland 1973; South 1976). Due to the expense and introduction of weed seed, the use of straw mulches has declined over time. Several managers were using pine straw in 1980 (Boyer and South 1984), but today few use it due to the expense. New bark or sawdust mulches are relatively weed-free (Stringfield 2005) but old, stockpiled supplies are often contaminated with weed seed. Several hardwood managers apply weed-free soil stabilizers after sowing. Most of these will forgo using mulch and therefore will apply additional irrigation.

Organic Amendments

In some cases, use of organic amendments will introduce weed seeds. In one nursery, rush (*Juncus* spp.) was introduced when an organic amendment was donated to the nursery. Yard litter and leaves collected by municipalities can contain many types of weed seeds. The value of these “free” amendments will depend on the increase in cost of subsequent weed control. Composting can help reduce the viability of many weed seeds but some will likely remain viable.

Handweeding

Frequent weeding can be an important IWM tool. Handweeding is best conducted when the soil is moist and weeds are small (figure 1). Weeding small plants has two advantages: the weeds are often removed before they go to seed, and the weeds are easier to remove when the roots are small. In many cases, the total weeding cost is less than if weeding is delayed until the weeds are large and hard to remove.

The use of seasonal labor varies with each nursery. When using contract labor, the cost of 100 hours of handweeding might exceed \$2000/acre. Therefore, the use of herbicides depends, in part, on the cost of handweeding. At some nurseries, herbicides are used and minimal handweeding is required while other managers rely on handweeding and, except for soil fumigants, do not apply herbicides to hardwood seedbeds. With an effective IWM program, an acre of hardwood seedbeds may require less than 25 hours of handweeding (South 2009).

Mechanical Cultivation

Mechanical cultivation for weed control between seedling rows is feasible when the spacing between rows is 12 in (30 cm) or wider (Stanley 1970; Barham 1980). Several types of seedbed and

alleyway cultivators are available (Lowman et al. 1992). For example, a “brush-hoe” can be effective in reducing weeds in hardwood seedbeds (South 1988) although there are some drawbacks. To obtain a specified level of weed control requires a precise adjustment to ensure a proper working depth (Weber 1994). Weeds within the row remain uninjured. Any small error in alignment can damage hardwood seedling roots or shoots. In 2006, only 2 hardwood managers (out of 26) were using mechanical weed control between seedling rows (South 2009).

Living Mulch

The “living mulch” concept was used by the Virginia Department of Forestry during the 1980s. Rye (*Secale cereale* L.) seed were drilled into the sections immediately before sowing hardwoods in the fall. The “living mulch” protected the fall sown seedbeds from injury by wind, rain, and frost. This system was also effective for fall-sown hardwoods in Illinois and Indiana (Stauder 1994, Wichman 1994). Nursery managers in Georgia and Tennessee currently sow wheat (*Triticum aestivum* L.), rye, or oats (*Avena sativa* L.) on prepared beds before fall sowing acorns (Ensminger 2002). The living mulch is then sprayed with an herbicide in February prior to emergence of oak seedlings. This system provides several advantages including a retardation of weed growth.

Fall Sowing

Fall-sown hardwoods, such as red oaks (*Quercus* spp.) and black walnut (*Juglans nigra* L.), typically have fewer weeds the following year than spring-sown crops. This reduction in weeds is due to application of herbicides sooner in the spring and the fact that fall-sown crops typically achieve full canopy closure and shade out weeds sooner than spring-sown crops (figure 2).

Soil Fumigation

At many nurseries, effective soil fumigation with methyl bromide has been a cornerstone of a successful IWM plan. Several nursery managers contend that soil fumigation is more important when growing hardwoods because there are fewer effective, registered herbicides than exist for conifer seedbeds (Murray 2009). It is relatively easy to justify soil fumigation because it typically costs less than 6 percent of the wholesale value of the hardwood crop. For this reason, most managers in the South fumigate the soil prior to each hardwood seedling crop. Although dazomet is used in northern hardwood nurseries (Schroeder and Alspach 1995, Storandt 2003), hardwood managers in the South have traditionally relied on a combination of methyl bromide and chloropicrin to reduce weed, nematode, and fungi populations.

In the future, methyl bromide will continue to be produced by oceans, fires, and certain plants and fungi. However, it is possible that it will no longer be used as a fumigant due to restrictions on production. If this occurs, some managers will likely switch to fumigants, such as chloropicrin and dazomet, that have relatively low efficacy on weeds. Although dazomet can control certain soilborne pests, it is not effective in controlling nutsedge (Carey 1995; Carey and South 1999; Fraedrich and Dwinell 2003). If the use of effective soil fumigants declines, nursery managers will need to increase herbicide use to control weeds in fallow fields or cover-crops.

Herbicides Use

The Weed Science Society of America (WSSA) sorts herbicides into 27 groups according to their chemical structure and activity. About one-third of these groups are used operationally in hardwood seedbeds (table 2). Herbicides in the cyclohexanedione family (WSSA group 1) and dinitroaniline family (WSSA group 3) are commonly used in hardwood seedbeds.

Herbicides can be grouped into selective (not generally harmful to hardwood seedlings) or nonselective (should not contact bark and foliage). Glyphosate is typically a nonselective herbicide (kills both weeds and hardwoods) while sethoxydim is a selective herbicide (kills only grasses) (South and Gjerstad 1982). It is important to know the specific crop/weed system involved. For example, the herbicide clopyralid is a selective herbicide for black walnut but it is nonselective when applied to black locust (*Robinia pseudoacacia* L.).

The terms preemergence or postemergence are used to describe when the herbicide is applied. For example, preemergence herbicides such as napropamide kill germinating weeds before they emerge through the soil surface. Some preemergence herbicides can be applied after emergence of the hardwood crop but before the emergence of the weed. Postemergence herbicides, on the other hand, are applied after the weeds emerge. When discussing herbicides, it is important to clarify if the application is to be made after the crop emerges and before the weeds emerge (e.g. pendimethalin, preemergence herbicide) or after weeds emerge but before the hardwoods emerge (e.g. glyphosate, postemergence herbicide).

Herbicide Applications in Cover Crops

The number of mature weeds in this year's cover crop will determine the amount of weed seeds present in next year's seedbeds. Some cover crops grow quickly and shade out the soil, thus reducing germination and growth of weeds. These cover crops are preferred over those that are sown at low densities and allow light to reach the soil. In the South, most herbicides used in cover crops will have no effect upon seedling growth the following year. This is especially true when the herbicide is applied before July 1. However, it is best to check with nursery experts to ensure that carryover from one season to the next will not be a problem. Some herbicide labels include information on the number of months required before sowing sensitive crops.

Cover-crop rotation provides an excellent opportunity to control weeds that are resistant to herbicides used in seedbeds. For example, if only diphenylether herbicides (WSSA group 14) were continually used on an area, resistant weed species such as prostrate spurge (*Euphorbia maculate* L.) could rapidly increase. However, by using an herbicide from a different herbicide family in the cover-crop area, the spread of troublesome weeds could be checked. Recommendations for using herbicides in cover crops vary depending upon the region and weed species to be controlled. Specific recommendations on herbicides and rates used can be obtained from the local extension service. Some genetically modified cover crops have a glyphosate-resistant gene that some managers use as part of an IWM program to reduce nutsedge in cover-crops.

Herbicide Applications on Fallow Land

Weed control with herbicides is much easier on fallow ground (figure 3) than it is in hardwood seedling beds because: (1) a greater number of herbicides may be applied to fallow ground; (2) injury from drift is less likely; (3) multiple applications can be made; (4) timing of the application is not restricted to stage of hardwood growth; and (5) it is easier to see the weeds. For troublesome weeds like nutsedge, use of multiple applications of glyphosate on fallow ground is the preferred method to reduce the number of tubers in the soil (Fraedrich et al. 2003). At some nurseries, more glyphosate is used in fallow fields than is used in bareroot seedbeds (Juntunen 2001).

Herbicide Applications on Riserlines and Fencerows

It is important to control weeds on riserlines and fencerows, not only to prevent weeds from producing seed, but also to reduce the cover available for small rodents. Some managers apply a tank mix of two or three preemergence herbicides to riserlines at the time of sowing to prevent weeds from maturing and going to seed. In contrast, a few managers will wait for weeds to develop and then apply a postemergence herbicide to kill emerged weeds. This type of application is often done with a shield designed to reduce drift to the hardwood crop (figure 4). Several types of shields can be used to reduce the potential of drift when applying herbicides to riserlines (Kees 2008). The number of herbicides that may be applied on riserlines is greater than the number the Environmental Protection Agency (EPA) allows to be applied to tree seedlings. To reduce the risk of injury, managers should avoid applying herbicides that are very water soluble (i.e. will move into adjacent seedbeds) or very persistent in the soil.

Herbicides Applications in Seedbeds

At Time of Sowing

Several hardwood nursery managers do not apply herbicides at time of sowing since they typically sow on recently fumigated fields. However, fumigated soils can easily be contaminated with wind-blown seed and therefore some managers apply herbicides at time of sowing (Jacob 2009, Murray 2009). Several preemergence herbicides can be applied at sowing to large-seeded species like oaks, black walnut, pecan (*Carya illinoensis* L.), and hickory (*Carya* spp.). In contrast, only a few preemergence herbicides may be applied to small-seeded species like American sycamore (*Platanus occidentalis* L.). Managers who apply herbicides at time of sowing generally have less weeding times than those who rely solely on soil fumigants.

Oxyflurofen is labeled for use on field-grown deciduous trees and has been used operationally as a preemergence herbicide (applied just after sowing) on large-seeded hardwoods (Jacob 2009, Murray 2009). Application should be made before seeds germinate since injury might occur if the herbicide contacts newly emerged tissues. When using oxyflurofen, large-seeded hardwoods can usually penetrate the herbicide barrier without much damage.

After the First True Leaves Have Formed

Herbicide selectivity is based on physiological or morphological differences between crop and weed. For example, a physiological difference between broadleaves and grasses is the basis of selectivity for clethodim, sethoxydim and fluziflop-butyl. As a result, these postemergence

herbicides typically do not cause injury to hardwoods after their first true leaves have formed. Preemergence herbicides (like prodiamine and pendimethalin) are active mainly on seed germination. These herbicides can also be applied once hardwood seedlings have germinated and have developed a few true leaves. The herbicide prodiamine is toxic to small hardwood seed such as sycamore if applied at time of seeding, but when applied after the seedlings are 2 in (5 cm) or taller, the chance of injury is greatly reduced. Although these herbicides will not control emerged weeds, they will help keep subsequent weed seed from germinating. This technique is used successfully by several nursery managers in the South.

Some foliar-acting postemergence herbicides (like clopyralid) are selective and will affect the foliage of some weeds without harming certain hardwoods (Lawrie and Clay 1994; Jacob 2009). However, clopyralid does have activity on legumes and therefore will injure eastern redbud (*Cercis canadensis* L.) and black locust. Injury has also been observed on black alder (*Alnus glutinosa* L.), hackberry (*Celtis occidentalis* L.), and dogwood (*Cornus florida* L.).

Granular Herbicide Formulations

The WSSA definition of “granular” is “a dry formulation consisting of discrete particles generally less than 10 mm³ and designed to be applied without a liquid carrier.” Granular herbicides are often used in horticultural nurseries and a number of granular herbicides are labeled for use on hardwoods. Although a number of granular herbicides may be used in hardwood nurseries, the cost is greater than for liquid formulations. The cost to treat an acre with granular herbicides could exceed \$120. This might be 8 to 10 times the cost of applying the same active ingredient sold as a liquid formulation.

An advantage of granular herbicides is that when hardwood leaves are dry, the granules drop to the ground and do not affect the foliage (figure 5). When applied to dry foliage, herbicide granules of oxyfluorfen and oxadiazon may be less phytotoxic to foliage than liquid formulations (which may contain inert ingredients like naphthalene). In cases where granules are lodged in the foliage, a sufficient amount of irrigation soon after treatment will reduce the chance of phototoxicity. For this reason, a wide variety of species are listed on granular herbicide labels. Granules of flumioxazin, oxyfluorfen, or oxadiazon could cause some temporary necrosis if they are allowed to remain on leaves.

Granular herbicides are not applied at time of sowing, but are applied after the hardwoods have developed true leaves. Although effective weed control can be obtained with granular herbicides (Reeder et al. 1991), some nursery managers choose not to use granular formulations due to the added expense and because application is easier when herbicides are sprayed.

Managers should be aware that “water dispersible granules” (WDG) do not fit the WSSA definition even though they are called “granules.” Therefore, do not treat WDG formulations as though they were true granular formulations. WDG formulations should be mixed with water and applied as a liquid spray. Do not apply WDG formulations without following the label directions.

Directed Herbicide Application Using Shields

One way to provide selectivity is to ensure the herbicide does not come in contact with the hardwood foliage. This can be accomplished with careful directed applications by hand or by using shields when applying herbicides between drill rows (figure 6). To reduce the potential for injury, most foliar-active herbicides should be directed away from the crop and toward the weeds.

Some nursery managers apply glyphosate “as needed” to control troublesome perennial weeds between rows using shielded sprayers (South and Carey 1999; Stallard 2005; Windell 2006). Glyphosate is a foliar-applied, nonselective herbicide with no soil activity. Glyphosate is bound tightly to soil particles and is unlikely to move off-site. The relatively slow absorption of glyphosate into foliage causes efficacy to be reduced by rains within a couple of hours of application. There is no legal limit to the number of applications that may be applied in a year.

Herbicide Injury

Although many factors can injure seedlings, herbicides are often the first to be blamed. For example, herbicides have been blamed for injury caused by fertilizer. In order to reduce a misdiagnosis, nursery managers should leave a few untreated areas in the seedbeds (i.e. check plots). The size of the check plot can be relatively small (perhaps 43 ft² [4 m²] per acre). These check plots are not only useful for diagnosing herbicide injury (figure 7), but they also provide a useful demonstration of what seedbeds would look like without the use of herbicides.

In some cases, herbicide injury will be minor and ephemeral. In fact, some herbicides might initially cause injury but eventually produce stock that is larger than untreated controls with no injury symptoms (Reeder et al. 1994). Therefore, most hardwood managers are more concerned with treatments that cause an “economic” injury to their crop than they are with one that causes a “cosmetic” injury to leaves, especially when hardwoods drop their leaves prior to lifting.

Economic injury occurs when an herbicide treatment reduces crop value (e.g. when the number of shippable seedlings produced per acre is reduced). The problem is determining which herbicides reduce seed germination prior to operational use. In some cases, herbicide trials are designed in such a manner that even a 50 percent reduction in crop value would not be classified as “significant” injury (Garrett et al. 1991, South 1992). The low power of these experimental designs is due primarily to the high level of variability in many hardwood seedbeds.

Herbicide injury can result when the label instructions or precautions are not followed. It is also important to be sure the herbicide sprayer is properly calibrated. Without regular calibration, uniformity may decrease and risk of injury could increase. Additionally, it is wise to consult with nursery experts prior to applying the herbicide since new information may have occurred since the label was written. For example, some managers have observed injury to dogwood when a certain herbicide in WSSA group 1 was applied to newly emerged seedlings. This is because one brand contained 65 percent solvent naphtha and 7 percent naphthalene (which can injure new foliage when applied under high temperatures). Consultation with an expert might have prevented this type of injury since they likely would have suggested using similar products that contained low amounts of naphtha and naphthalene.

Occasionally, hardwoods have been injured when environmental conditions are right and the herbicide “lifts-off” the soil within water vapor and then drift over newly-emerged hardwood seedlings, a process known as co-distillation. This may occur soon after seedbeds have been treated with oxyfluorfen on warm, sunny days. The injury is usually just cosmetic as the new leaves turn brown. Fortunately, the affected seedlings typically recover and grow normally.

At a few nurseries, the use of dinitroaniline herbicides (WSSA group 3) has injured certain hardwood species (Warren and Skroch 1991, Hood and Klett 1992, South 1992, Derr and Salihu 1996). In some cases, herbicide galls formed on the stem near the groundline (Altland 2005, South 2009). For example, sugarberry (*Celtis laevigata* Willd.) was injured after applying prodiamine and pendimethalin (figure 7). A postemergence application (after both weed and crop emergence) of oryzalin has caused injury and stem breakage on American sycamore, river birch (*Betula nigra* L.), yellow poplar (*Liriodendron tulipifera* L.), redbud, elm (*Ulmus* spp.), buttonbush (*Cephalanthus* spp.), plum (*Prunus* spp.), and black willow (*Salix nigra* L.).

In some cases, herbicide injury occurs when an herbicide applied to fallow ground carries over to the next year. For example, injury occurred when certain herbicides in the imidazolinone family were used the previous year on fallow land. The length of time that an herbicide remains biologically active in the soil is determined by a number of factors. In the South, most herbicides in WSSA groups 1, 3, and 15 do not persist long enough to affect hardwoods sown the next year. In regions where soils are cooler (e.g. Saskatchewan), however, herbicide carryover is more likely. This is primarily because the rate of microbial decomposition is slower in Saskatchewan than in Alabama or Georgia.

Occasionally, herbicide injury will occur after a wind storm. For example, at one nursery, herbicide injury to Sawtooth oak (*Quercus acutissima* L.), swamp chestnut oak (*Quercus michauxii* Nutt.), and persimmon (*Diospyros virginiana* L.) was noticed two days after seedlings were sandblasted (Skidmore 1966) with high winds. The herbicide (i.e. oxyfluorfen), was carried with the soil and the abrasions allowed the herbicide to enter the stem and foliage. Although oak seedlings in check plots were also sandblasted (figure 7), they were not injured since the sand did not contain the herbicide. Use of a soil stabilizer would have reduced the amount of sandblasting and this would have subsequently reduced this type of injury.

In some situations, injury to adjacent seedbeds has occurred when dazomet or metham sodium was applied without a tarp (Scholtes 1988, Buzzo 2003, Starkey 2011). Therefore, to reduce the potential for injury to adjacent crops, a plastic tarp is recommended when fumigating with these compounds. Some contractors now only use a plastic tarp when fumigating with metham sodium.

When using liquid fertilizers in returnable totes, it is wise to deal with a reputable dealer. Reputable dealers either do not refill herbicide totes with fertilizer solutions or they ensure the totes are thoroughly cleaned before they are refilled. At one nursery, injury resulted when a fertilizer dealer did not thoroughly clean out a tote that had previously contained triclopyr.

Economics

Some nursery managers base their weed management decisions on securing economic profits and on maintaining good reputations for producing high-quality nursery stock. Their justifications for using herbicides include keeping seed efficiency high (South 1987) and production costs low. In contrast, other managers operate as a non-profit entity and their primary objective is to achieve target production goals within a given budget. Both management systems can benefit when handweeding costs are reduced by using an effective IWM program.

The easiest way to justify the use of herbicides is to compare the cost of treatment with the cost of handweeding. For example, at a nursery where hand labor costs \$15/hr, an herbicide application that costs \$30/acre would be justified if it reduced handweeding times by 2 hr/ac or more. Thus, if a nursery required 125 hr/ac of handweeding (\$1,875 total) to remove small grasses, ten applications of herbicides (i.e. \$300/ac) could reduce weed control costs by as much as \$1,575/ac (assuming the use of herbicides eliminated the need for handweeding the grass).

Another method to justify herbicide use is to determine how many seedlings are lost to weed competition and to handweeding. If a nursery loses \$0.30 every time a seedling is inadvertently pulled up by a weeder, then saving 100 seedlings could justify an herbicide treatment that cost \$30/acre. Therefore, even in rare cases where use of herbicides does not reduce the annual cost of weed control, their use could still be justified when seedling sales are increased. An examination of a hardwood nursery budget might reveal that herbicide treatments amount to less than 0.5 percent of the retail value of the crop (table 3). Therefore, use of herbicides may be justified when seedling production (i.e. number of seedlings sold per acre) is increased by just 0.5 percent. This would be equivalent to selling 201,000 seedlings instead of 200,000 seedlings.

Summary

Due to the numerous species involved, a single herbicide regime (e.g. South 1992) is unlikely to be effective for all hardwood species. Weed species, hardwood species, soil types, and labor costs vary with nursery and therefore weed management regimes vary with nursery. However, the most effective IWM programs include a rigorous sanitation program and judicious use of efficacious herbicides.

Disclaimer:

The mention of commercial products is solely for the information of the reader. Endorsement is not intended.

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REFERENCES

- Altland, J.E. 2005. Weed control in nursery field production. Extension Service Report EM8899-E. Eugene, OR: Oregon State University.
http://oregonstate.edu/dept/nursery-weeds/feature_articles/em8899.pdf. (August 2015)
- Abrahamson, L.P. 1987. Forest tree nursery herbicide studies at the Oklahoma forest regeneration center. In: Landis, T.D., tech. coord. National proceedings, intermountain forest nursery association. General Technical Report RM-151. Fort Collins, CO: U.S. Department of Agriculture, Forest Service, Rocky Mountain Forest and Range Experiment Station: 49–57.
- Bland, W.A. 1973. Study to evaluate the effects and costs of mulching materials in loblolly pine seedbeds. Forestry Note No. 3. Raleigh, NC: North Carolina Forest Service.
- Barham, R.O. 1980. Handweeding times reduced in hardwood seedbeds by a modified rolling cultivator. Tree Planters' Notes 31(4):30-32.
- Boyer, J.N.; South, D.B. 1984. Forest nursery practices in the South. Southern Journal of Applied Forestry. 8(2): 67-75.
- Buzzo, R.J. 2003. Phytotoxicity with metam sodium. In: Riley L.E., Dumroese R.K., Landis T.D., tech. coords. National proceedings: forest and conservation nursery associations – 2002. Proceedings RMRS-P-28. Ogden, UT: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station: 79–83
- Carey, W.A. 1995. Chemical alternatives to methyl bromide. In: Landis, T.D.; Dumroese, R.K., tech. coords. National proceedings: forest and conservation nursery associations – 1994. General Technical Report RM-257. Fort Collins, CO: U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station: 4-11.
- Carey, W.A.; South, D.B. 1999. Effect of chloropicrin, vapam, and herbicides for the control of purple nutsedge in southern pine nurseries. In: Landis, T.D.; Barnett, J.P., tech. coords. National proceedings: forest and conservation nursery associations – 1998. General Technical Report SRS-25. Asheville, NC: U.S. Department of Agriculture, Southern Research Station: 39-40.
- Derr, J.F.; Salihu, S. 1996. Preemergence herbicide effects on nursery crop root and shoot growth. Journal of Environmental Horticulture. 14(4):210-213.
- Ensminger, P. 2002. Nursery practices in Tennessee. In: Dumrose, R.K.; Riley, L.E.; Landis, T.D., tech. coords. National proceedings: forest and conservation nursery associations – 1999, 2000, and 2001. Proceedings RMRS-P-24. Ogden, UT: U.S. Department of Agriculture, Rocky Mountain Research Station: 281-283.
- Fraedrich, S.W.; Dwindell, L.D. 2003. An evaluation of dazomet incorporation methods on soilborne organisms and pine seedling production in southern nurseries. Southern Journal of Applied Forestry. 27(1):41-51.

Fraedrich, S.W.; Dwindell, L.D.; Cram M.M. 2003. Broadcast applications of glyphosate control nutsedge at a south Georgia forest tree nursery. *Southern Journal of Applied Forestry*. 27(3):176-179.

Garrett, H.E.; Stenberg, R.C.; Cox, G.S.; Mitchell, R.J.; Young, W.G. 1991. A case for herbicidal weed control in forest nurseries. In: Kaufman, J.E.; Westerdahl, H.E., eds. *Chemical Vegetation Management*. Athens, GA.: Plant Growth Regulator Society of America: 83-101. Chapter 7.

Hood, L.R.; Klett, J.E. 1992. Preemergent weed control in container-grown herbaceous and woody plants. *Journal of environmental Horticulture*. 10(1):8-11.

Ishii, K.; Yamagi, K.; Manabe, T. 1971. Development and chemical control of purple nutsedge in the forest nursery. *Weed Research, Japan*. 12: 45-49.

Jacob, R. 2009. Hardwood weed control: Iowa Department of Natural Resources Forestry, Iowa State Nursery. In: Dumroese, R.K.; Riley, L.E., tech. coords. *National proceedings: forest and conservation nursery associations -2008*. Proceedings RMRS-P-58. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 76-78.

Juntunen, M.L. 2001. Use of pesticides in Finnish forest nurseries in 1996. *Silva Fennica*. 35(2):147-157.

Kees, G. 2008. Herbicide shield for spraying irrigation pipelines. Tech Tip 0824-2343-MTDC. Missoula, MT: USDA Forest Service, Missoula Technology and Development Center. 4 p. <http://www.fs.fed.us/t-d/pubs/pdfpubs/pdf08242343/pdf08242343dpi300.pdf>. (August 2015)

Klingman, G.C.; Ashton, F.M. 1975. *Weed science: Principles and practices*. NY: John Wiley and Sons. 431 p.

Lawrie, J.; Clay, D.V. 1994. Tolerance of 2-year-old forestry trees to five herbicides. *Forestry*. 67(4): 287-295.

Lowman, B.j.; Landis, T.D.; Zensen, F.; Holland, B.J. 1992. Bareroot Nursery Equipment Catalog. MTDC Project Report No. 9224-2839-MTDC. Missoula, MT: USDA Forest Service, Missoula technology and Development Center: 198.

Mullin, R.E. 1965. Effects of mulches on nursery seedbeds of white spruce. *The Forestry Chronicle*. 41(4): 454-465.

Murray, A. 2009. Successes and failures in controlling weeds in hardwood seedbeds at the Arkansas Forestry Commission Baucum Forest Nursery. In: Dumroese, R.K.; Riley, L.E., tech. coords. *National proceedings: forest and conservation nursery associations -2008*. Proceedings RMRS-P-58. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 74-75.

Reeder, J.A.; Gilliam, C.H.; Wehtje, G.R.; South, D.B. 1991. The effects of selected herbicides on propagation of chestnut oaks in containers. Combined Proceedings of the International Plant Propagators' Society. 41:325-329.

Reeder, J.A.; Gilliam, C.H.; Wehtje, G.R.; South, D.B.; Keever, G.J. 1994. Evaluation of selected herbicides on field-grown woody ornamentals. Journal of Environmental Horticulture. 12(4):236-240.

Scholtes, J.R. 1989. Soil fumigation at J. Herbert Stone Nursery. In: Landis, T.D., tech. coord. National proceedings: intermountain forest nursery association – 1989. General Technical Report RM-184. Fort Collins, CO: U.S. Department of Agriculture, Pacific Northwest Research Station: 35-37.

Schroeder, W.R.; Alspach, L.K. 1995. Herbicide program at the PFRA shelterbelt centre. In: Landis, T.D., Cregg, B., tech. coord. National proceedings: forest and conservation nursery associations – 1995. General Technical Report PNW-365. Portland, OR: U.S. Department of Agriculture, Pacific Northwest Research Station: 80-83.

Skidmore, E.L. 1966. Wind and sandblast injury to seedling green beans. Agronomy Journal. 58:311-315.

South, D. 1976. Pine straw mulch increases weeds in forest tree nurseries. Agricultural Experiment Station, Auburn University. Highlights of Agricultural Research. 23(4):15.

South, D.B. 1984. Response of loblolly pine and sweetgum seedlings to oxyfluorfen. Canadian Journal of Forest Research. 14(4):610-604.

South, D.B. 1984. Chemical weed control in southern hardwood nurseries. Southern Journal of Applied Forestry. 8(1):16-22.

South, D.B. 1987. Economic aspects of nursery seed efficiency. Southern Journal of Applied Forestry. 11(2):106-109.

South, D.B. 1988. Mechanical weed control for the forest nursery. Georgia Forestry Commission Research Report. 1: 1-9.

South, D.B. 1992. Prodiamine: a herbicide for pine and hardwood nurseries. Southern Journal of Applied Forestry. 16(3):142-146.

South, D.B. 2000. Tolerance of southern pine seedlings to clopyralid. Southern Journal of Applied Forestry. 24(1):51-56.

South, D.B. 2009. A century of progress in weed control in hardwood seedbeds. In: Dumroese, R.K.; Riley, L.E., tech. coords. National proceedings: forest and conservation nursery associations –2008. Proceedings RMRS-P-58. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 80-84.

South, D.B.; Carey, W.A. 2005. Weed control in bareroot hardwood nurseries. In: Dumroese, R.K.; Riley, L.E.; Landis, T.D., tech. coords. National proceedings: forest and conservation nursery associations – 2004. Proceedings RMRS-P-35. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 34-38.

South, D.B.; Gjerstad, D.H. 1982. Postemergence control of grasses with selective herbicides in pine and hardwood seedbeds. *Tree Planters' Notes*. 33(1): 24-28.

Stauder, A.F. 1994. The use of green overwinter mulch in the Illinois State Nursery program. In: Landis, T.D., tech. coord. National proceedings, northeastern and intermountain forest and conservation nursery association –1993. General Technical Report RM-243. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Forest and Range Experiment Station: 51-53.

Stallard, D.H. 2005. Panel Discussion: Using Shielded Sprayers to Control Weeds in Nursery Beds. In: Dumroese, R.K.; Riley, L.E.; Landis, T.D., tech. coords. National proceedings: forest and conservation nursery associations – 2004. Proceedings RMRS-P-35. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 24-25

Stanley, H. 1970. Hardwood weed control. In: Jones, L., tech coord. Proceedings, southeastern nurserymen's conferences. Atlanta, GA: U.S. Department of Agriculture, Forest Service, Southeastern Area: 60-61.

Starkey, T.E. 2011. The history and future of methyl bromide alternatives in the southern United States. In: Haase, D.L., Pinto, J.R. Riley, L.E., tech. coords. National proceedings: forest and conservation nursery associations —2011. Proceedings RMRS-P-68. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 31-35.

Stevens, O.A. 1932. The number and weight of seeds produced by weeds. *American Journal of Botany*. 19(9): 784-794.

Storandt, J. 2002. Red oak propagation at the Griffith State Nursery, Wisconsin Rapids, Wisconsin. In: Dumroese, R.K.; Riley, L.E.; Landis T.D., tech. coords. National proceedings: forest and conservation nursery associations –1999, 2000, and 2001. Proceedings RMRS-P-24. Ogden, UT; U.S. Department of Agriculture, Rocky Mountain Research Station: 120-121.

Stringfield, D. 2005. Panel discussion: weed management. In: Dumroese, R.K.; Riley, L.E.; Landis, T.D., tech. coords. National proceedings: forest and conservation nursery associations –2004. Proceedings RMRS-P-35. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Research Station: 39-40.

Walker, R.H.; Buchanan, G.A. 1982. Crop manipulation in integrated weed management systems. *Weed Science*. 30 (Suppl. 1): 17-24.

Warren, S.L.; Skroch, W.A. 1991. Evaluation of six herbicides for potential use in tree seedbeds. *Journal of Environmental Horticulture*. 9(3):160-163.

Weber, H. 1994. Mechanical weed control with a row brush hoe. *Acta Horticulturae*. 372: 253- 260

Wichman, J.R. 1982. Weed sanitation program at the Vallonia Nursery. *Tree Planters' Notes*. 33(4):35-36.

Wichman, J.R. 1994. Use of wheat as a living mulch to replace hydromulch for fall sown seedbeds. In: Landis, T.D., tech. coord. National Proceedings, northeastern and intermountain forest and conservation nursery association – 1993. General Technical Report RM-243. Fort Collins, CO: U.S. Department of Agriculture, Rocky Mountain Forest and Range Experiment Station: 55-56.

Windell, K. 2006. Shielded herbicide sprayer for hardwood nursery seedling beds. Technical Report 0624-2827-MTDC. Missoula, MT: U.S. Department of Agriculture, Missoula Technology and Development Center. 18 p.
<http://www.fs.fed.us/t-d/pubs/pdfpubs/pdf06242827/pdf06242827dpi300.pdf>. (August 2015)

Table 1. Typical weed species in southern hardwood nurseries.

Common name	Scientific name
Grasses	
bermudagrass	<i>Cynodon dactylon</i> (L.) Pers.
crowfootgrass	<i>Dactyloctenium aegyptium</i> (L.) Richter
hairy crabgrass	<i>Digitaria sanguinalis</i> (L.) Scop.
sourgrass	<i>Digitaria insularis</i> (L.) Mez ex Ekman
barnyardgrass	<i>Echinochloa crus-galli</i> (L.) Beauv.
goosegrass	<i>Elusine indica</i> (L.) Gaertn.
Sedges	
flathead sedge	<i>Cyperus compressus</i> L.
yellow nutsedge	<i>Cyperus esculentus</i> L.
purple nutsedge	<i>Cyperus rotundus</i> L.
Broadleaves	
prostrate pigweed	<i>Amaranthus bitoides</i> S. Wats.
redroot pigweed	<i>Amaranthus retroflexus</i> L.
spiny amaranth	<i>Amaranthus spinosus</i> L.
sicklepod	<i>Cassia obtusifolia</i> L.
eclipta	<i>Eclipta alba</i> (L.) Hassk.
dogfennel	<i>Euportorium capillifolium</i> (Lam.) Small
spurge	<i>Chamaesyce maculata</i> (L.) Small
tall morning glory	<i>Ipomoea purpurea</i> (L.) Roth
carpetweed	<i>Mollugo verticillata</i> L.
white clover	<i>Trifolium repens</i> L.

Table 2. Common names and trade names of selected herbicides used in southern hardwood nurseries.

Common Name	Product Names	Comment	WSSA group	REI* hours
Soil Fumigant				
chloropicrin	Various	Good nutsedge control	n/a	>120
dazomet	Basamid	Poor nutsedge control	27	>120
methyl bromide	Various	Excellent nutsedge control	n/a	>120
After sowing for oaks, walnut, hickory				
oxyfluorfen	Goal, Galigan, Goaltender	Field-grown	14	24
Very selective grass herbicides				
clethodim	Clethodim, Select, Shadow	Grass control only	1	24
fluzafop	Fusilade	Grass control only	1	12
sethoxydim	Segment, Sethoxydim	Grass control only	1	12
Herbicides with some selectivity when applied over established hardwoods				
DCPA	Dacthal	Found in groundwater	3	12
dithiopyr	Dimension	Established plants only	3	12
oryzalin	Surflan	May cause galls	3	24
pendimethalin	Pendulum (Aquacap)	May cause galls	3	24
prodiamine	Barricade	May cause galls	3	12
trifluralin	Trifluralin HF	Certain labels only	3	12
clopyralid	Lontrel	Will injure legumes	4	12
oxyfluorfen	Goaltender	Field-grown	14	24
S-metolachlor	Pennant	Active on sedge	15	24
napropamide	Devrinol	Some grass control	15	12
Granules Herbicides – can be applied over transplanted stock				
flumioxazin	Broadstar	Apply to dry leaves Do not apply to bedding plants	14	12
oxadiazon	Ronstar	Apply to dry leaves	14	12
oxyfluorfen + pendimethalin	OH2	Apply to dry leaves	14 3	24
dimethanamid + pendimethalin	Freehand	May cause galls	15 3	24
To avoid injury, do not apply to seedbeds				
dichlobenil	Casoron	4 weeks after transplanting	20	12
pronamide	Kerb	Not for use on 1-0 stock	3	24
isoxaben	Gallery	Assume all risks	21	12
simazine	Princep	Oak transplants	5	12
Nonselective Herbicides – applications must be directed away from seedlings				
glyphosate	Roundup	Use shielded applicator	9	4
pelargonic acid	Scythe	Use shielded applicator	27	12
sulfosulfuron	Certainty	Avoid contact with leaves	2	12

*REI: Restricted-entry intervals for agricultural uses.

Check the AGRICULTURAL USE REQUIREMENTS section of the label for required REI.

Table 3. Example of weed management costs in southern hardwood nurseries. Data assumes 180,000 seedlings/ac and a price of \$0.30/seedling.

Weed management practice	Active ingredient lbs/acre	\$ Per acre	\$ Per thousand seedlings	Percentage of total crop value
Herbicides in seedbeds	2	90	\$0.50	0.2%
Herbicides on fallow ground	--	--	\$0.70	0.2%
Handweeding – 25 hours	0	375	\$2.08	0.7%
Soil fumigation	350	3,100	\$17.22	5.7%
Total			\$20.50	6.8%



Figure 1. When nursery managers adopt an effective integrated weed management program, the amount of handweeding can be kept to less than 25 hours per acre. Hand weeding is most effective when weeds are small, before they go to seed. Weeding takes less time when the soil is moist and the weed has a small root system. (Photo by Sam Campbell 2010)



Figure 2. Weed control is typically easier when hardwoods are sown in the fall or winter. This is because the canopy closes sooner in the spring and the resulting shade reduces growth of various weed species. (Photo by Don Stringfield 2002)



Figure 3. An effective way to control nutsedge of fallow ground is to treat emerged plants with glyphosate. Some managers treat nutsedge two or three times from June to September to reduce the population of tubers in the soil. (Photo by David South 2005)



Figure 4. Controlling weeds adjacent to seedbeds is an important part of an integrated weed management program. Some managers prefer to apply preemergence herbicides to irrigation lines at time of sowing hardwoods while others wait to treat emerged weeds with postemergence herbicides that have no soil activity. (Photo by Keith Windell 2001)



Figure 5. Although granular herbicides are typically more expensive, they often are less phytotoxic than liquid formulations. Granular herbicides should be applied to dry foliage which allows most of the granules to roll off the foliage. Those that remain lodged in the foliage could be shaken off by dragging a cloth or bar over the foliage. (Photo by David South 2007)



Figure 6. Pictured is an example of a shielded herbicide applicator designed for hardwood seedbeds. This model has an advantage in that it can be operated by one person (Photo by David South 2010)



Figure 7. Use of untreated check plots can help to properly identify herbicide injury. Seedlings on both seedbeds experienced sandblasting during a May storm. Seedlings on the bed on the left were injured by sand that carried an herbicide while those on the right were blasted with soil that did not contain a herbicide. In this case, injury was temporary and seedlings were fully recovered by July. (Photo by David South 2010)



Figure 8. Some hardwood species are more tolerant of herbicides than others. For example, sugarberry (*Celtis laevigata* Willdenow) can be injured by certain dinitroaniline herbicides. (Photo by Chase Weatherby 2008)